

计算光学成像与光信息处理技术前沿 第十二周课(lecture 8)

差分相衬成像

左超

南京理工大学电光学院光电技术系

Jiangsu Key Laboratory of Spectral Imaging & Intelligent Sense (SIIS) Nanjing University of Science and Technology, Nanjing, Jiangsu Province 210094, China











振幅

□ 成像依赖染色标记 □ 无法定量探测相位







无标记成像 (Label-free)







Staining and fluorescence (a)



Fluorescence techniques

Confocal microscopy; Total internal reflection fluorescence; Two/multi-photon microscopy; Light-sheet microscopy;

. . .

Super-resolution fluorescence techniques

. . .

Stimulated emission depletion microscopy; Photoactivated localization microscopy; Stochastic optical reconstruction microscopy; Structured illumination microscopy;



Noninterferometic QPI techniques

. . .

Laser

Transport- of-intensity equation; Differential phase contrast; Fourier ptychographic microscopy; Lens-free on-chip holography;

Optical interferometry and holography

Digital holographic microscopy

Phase contrast techniques

Zernike phase-contrast microscopy Differential interference contrast microscopy Computational light microscopy

CCD

(b) Phase changes

"无标记"相衬成像技术









Dark field and phase contrast microscopes



b

P

20

1C)



Phase contrast

INSTITUTE OF SMALL WORLD

传统显微成像

计算光学显微成像



	Source	····▶ [%] ····· Sampl e	Imaging system	O O O O O O O O O O O O O O O O O
Principle	Linear decomposition	Iteraction Diffraction theory	Propagation or modulation	Intensity detection and sampling
Solution	Source points coherent modes	Scalar diffraction theory; Vector diffraction theory;	Linear systems theory	Amplitude square Non-coherent superposition
Analysis	Coherent properties	Superposition under plane-wave illumination	Linear spatially invariant system;	Time average
Model	Complex intensity; Wavelengh; Coherence function; Spectrum distribution; Azimuth; Polarization	Complex transmittance function; Absorption function; Phase function; Refractive index; Polarization characteristics;	2D/3D coherent spread function 2D/3D point spread function; Coherent transfer function; Optical transfer function; 4D phase-space model;	Spectral response function; Quantum efficiency; Single Photon; Noise statistics; Modulation transfer function; Nonlinearity;
Modulation	Intensity; Phase; Angle; Aperture; Wavelength; Spatial coherence; Temporal coherence; Polarization; Paraxial approximation; etc.	Pure phase/weak object/ slowly varying approximations; Born or Rytvo approximation; Multi-slice propagation model; Non-negative refractive index approximation; etc.	Imaging aperture; Out-of-focus; Light deflection; Light intensity masks; Chromatic dispersion etc.	Controlled displacement: lateral and axial; Spectrum integration: spectrum response color cross-talk martix; Pixelation effects: binning, downsampling, pixel response functon, Bayer filter; Polarization detection; etc.



□ 倾斜照明





□ 倾斜照明

$$F_{w} = F_{ae^{i\phi}} = W$$
 傅立叶变换对
 $F_{we^{i\theta}} = F_{ae^{i\phi}e^{i\theta}} = W(u - k_x, v - k_y)$

 a
 $\varphi + \theta$
 $W(u - k_x, v - k_y)$

 Image: Comparison of the system of the sys



□ 倾斜照明





口部分相干成像系统图像生成模型

强度分布:

$$I(\mathbf{x}) = \iiint S(\mathbf{u})\hat{T}(\mathbf{u}_1)\hat{T}^*(\mathbf{u}_2)H(\mathbf{u}+\mathbf{u}_1)H^*(\mathbf{u}+\mathbf{u}_2)e^{j2\pi\mathbf{x}(\mathbf{u}_1-\mathbf{u}_2)}d\mathbf{u}_1d\mathbf{u}_2d\mathbf{u$$

分离与样品**无关的系统**项:

$$I(\mathbf{x}) = \iint \hat{T}(\mathbf{u}_1) \hat{T}^*(\mathbf{u}_2) TCC(\mathbf{u}_1, \mathbf{u}_2) e^{j2\pi \mathbf{x}(\mathbf{u}_1 - \mathbf{u}_2)} d\mathbf{u}_1 d\mathbf{u}_2$$

部分相干传递函数/交叉传递系数

$$C(\mathbf{u}_1,\mathbf{u}_2) = \iint S(\mathbf{u})H(\mathbf{u}+\mathbf{u}_1)H^*(\mathbf{u}+\mathbf{u}_2)d\mathbf{u}$$



口部分相干成像系统图像生成模型

$$I(\mathbf{x}) = \iiint S(\mathbf{u})\hat{T}(\mathbf{u}_1)\hat{T}^*(\mathbf{u}_2)H(\mathbf{u}+\mathbf{u}_1)H^*(\mathbf{u}+\mathbf{u}_2)e^{j2\pi\mathbf{x}(\mathbf{u}_1-\mathbf{u}_2)}d\mathbf{u}_1d\mathbf{u}_2d\mathbf{u$$



Hamilton, D. K., C. J. R. Sheppard, and T. Wilson. "Improved imaging of phase gradients in scanning optical microscopy." Journal of microscopy 135.3 (1984): 275-286.



□弱物体**近**似

$$A(\mathbf{u}) = \iint S(\mathbf{u}') P^*(\mathbf{u}') P(\mathbf{u} + \mathbf{u}') d\mathbf{u}' + \iint S(\mathbf{u}') P(\mathbf{u}') P^*(\mathbf{u} - \mathbf{u}') d\mathbf{u}'$$
$$P(\mathbf{u}) = \iint S(\mathbf{u}') P^*(\mathbf{u}') P(\mathbf{u} + \mathbf{u}') d\mathbf{u}' - \iint S(\mathbf{u}') P(\mathbf{u}') P^*(\mathbf{u} - \mathbf{u}') d\mathbf{u}'$$





□ 弱物体近似下的定量相位求解

$$I(x) = B\delta(\mathbf{u}) + A(\mathbf{u})a(\mathbf{u}) + jP(\mathbf{u})\phi(\mathbf{u})$$



Hamilton, D. K., & Sheppard, C. J. R. (1984). Journal of microscopy, 133(1), 27-



□缓变物体近似



相位梯**度传**递函**数**

$$PGTH = C(\mathbf{u}, \mathbf{u}) = \int S(\mathbf{u}' - \mathbf{u}) |P(\mathbf{u}')|^2 d\mathbf{u}'$$

相位梯度积分

$$\phi = \operatorname{Im} \left[\mathcal{F}^{-1} \begin{cases} \frac{\mathcal{F} \left\{ \phi'_{x} + \phi'_{y} \right\}}{2\pi \left(u + iv \right)}, & |\mathbf{u}| \neq 0 \\ C, & |\mathbf{u}| = 0 \end{cases} \right]$$





扫描式差分相衬成像系统

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Differential phase contrast in scanning optical microscopy

by D. K. HAMILTON and C. J. R. SHEPPARD, University of Oxford, Department of Engineering Science, Parks Road, Oxford OX1 3PJ

KEY WORDS. Scanning optical microscopy, differential interference contrast, Nomarski optics, laser, biological specimens, image formation, high resolution, optical microscopy.

SUMMARY

High-quality high-resolution transmission and reflection images produced using a scanning optical microscope and the split-detector technique are presented. These images exhibit differential phase contrast, the method avoiding some drawbacks of the usual Nomarski DIC arrangement. Imaging is treated theoretically and compared with the Nomarski method.

1. INTRODUCTION

Although Nomarski differential interference contrast (Nomarski, 1955) has become a widely used and powerful technique in optical microscopy it does have a number of disadvantages. A compromise must be made between contrast and signal level so that for objects with weak variations the signal may be very weak and in order to obtain adequate contrast the condenser must often be stepped down somewhat so that optimum resolution is no longer achieved. In general the image is formed by a complicated mixture of different contrast mechanisms including non-differential amplitude and differential amplitude contrast and with birefringent specimens it must be used with care.

A method widely used for scanning transmission electron microscopy (Dekkers & de Lang, 1974, 1977) based on a split detector, overcomes these problems, and indeed its use in scanning optical microscopy was suggested in the paper in which it was originally described (Dekkers & de Lang, 1974). However, until now, production of high-quality high-resolution optical images using the technique has not been reported.

Hamilton, D. K., & Sheppard, C. J. R. (1984).













差分相衬成像技术发展



全场差分相衬成像系统

1924 OPTICS LETTERS / Vol. 34, No. 13 / July 1, 2009

Quantitative phase-gradient imaging at high resolution with asymmetric illumination-based differential phase contrast

Shalin B. Mehta^{1,2,4,*} and Colin J. R. Sheppard^{1,2,3}

¹Optical Bioimaging Lab, Division of Bioengineering, National University of Singapore, Block-E3A, #7-10, 7 Engineering Drive 1, Singapore 117574 ²NUS Graduate School for Integrative Sciences and Engineering, 28 Medical Drive, Singapore 117456 ³Department of Biological Sciences, National University of Singapore, 14 Science Drive, Singapore 117543 ⁴ shalin@nus.edu.sg *Corresponding author: shalin@nus.edu.sg

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We describe a full-field phase-gradient imaging method: asymmetric illumination-based differential phase contrast (AIDPC). Imaging properties of AIDPC are evaluated using the phase-gradient transfer-function approach and elucidated with experimental images of an optical fiber and a histochemical preparation of a skeletal muscle section. In comparison with full-field differential interference contrast, AIDPC does not require phase shifting for quantitative imaging of phase gradient, provides artifact-free images of birefringent specimens, requires shorter camera exposure, and has larger depth of focus. It is amenable to transfer-function engineering, simultaneous fluorescence imaging, and automated live cell imaging. © 2009 Optical Society of America

OCIS codes: 110.0180, 110.4850, 100.5070, 110.4980.

Imaging of biological specimens requires special optical processing to translate the optical thickness (i.e., phase) information to image intensity. Direct quantitative measurement of phase requires use of coherent illumination, leading to limited spatial resolution, lack of optical sectioning, and speckle from imperfections in the optical train. Phase-gradient imaging methods such as Nomarski's differential interference contrast (DIC) can accommodate large illumination apertures (i.e., partially coherent illumination), alleviating the above problems. In scanning optical microscopy, an intrinsically linear phase-gradient contrast method, termed differential phase contrast (DPC) has been evaluated [1-3]. In contrast to DPC, DIC images a complex mix of amplitude and phasegradient information, necessitating approaches such as phase shifting (PSDIC) to establish a linear rela-

tical to those of the full-field system with an incoherent source if two conditions are met [11]: (1) each system has the same objective apertures (i.e., pupil amplitude), $P_o(\xi, \eta)$ and (2) the sensitivity distribution of the detector in the scanning system is the same as the intensity distribution of the condenser aperture, $|P_c(\xi,\eta)|^2$, in the full-field system. In scanning DPC, a split-detector or a quadrant diode is placed in the Fourier plane of the collector, and the image is formed by subtracting intensities recorded by two halves of the detector. A reciprocal wide-field DPC system has an antisymmetric condenser aperture with half of the aperture having negative effective intensity. We synthesize negative condenser intensity by subtracting two images acquired with semicircular condenser apertures in the direction of differentiation



Mehta S. B. & Sheppard, C. J. R. (2009).



口基于LCD的可编程孔径差分相衬成像系统





口和内窥镜相结合,用于手术过程中成像









口结合智能手机,便携**化差分相**衬**成像系统**





Quantitative differential phase contrast imaging in an LED array microscope

Lei Tian^{1,*} and Laura Waller² ¹Department of Electrical Engineering and Computer Sciences, University of California, Berkeley, CA, 94709, USA *lei tian@alum mit edu

Abstract: Illumination-based differential phase contrast (DPC) is a phase imaging method that uses a pair of images with asymmetric illumination patterns. Distinct from coherent techniques, DPC relies on spatially partially coherent light, providing 2× better lateral resolution, better optical sectioning and immunity to speckle noise. In this paper, we derive the 2D weak object transfer function (WOTF) and develop a quantitative phase reconstruction method that is robust to noise. The effect of spatial coherence is studied experimentally, and multiple-angle DPC is shown to provide improved frequency coverage for more stable phase recovery. Our method uses an LED array microscope to achieve real-time (10 Hz) quantitative phase imaging with in vitro live cell samples.

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OCIS codes: (100.5070) Phase retrieval; (110.1758) Computational imaging; (170.0180) Microscopy; (110.3010) Image reconstruction techniques.

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Tian L., Waller L. (2015).









Step 2 Calculating the phase gradient Calculating the PTF Step 4 Step 3 Deconvolution Phase gradient Deconvolution Calculating phase gradient Calculating the PTF Step 1 S_L , S_R , S_U , S_D I_L , I_R , I_U , I_D Phase gradient Reconstructed phase







差分相衬定量相位成像



□频谱带宽受限,低频对比度差,无法恢复高分辨率细节
 □传递响应各向异性,成像分辨率各向异性





$$PTF(\mathbf{u}) = \frac{\iint S(\mathbf{u}_j) [P^*(\mathbf{u}_j) P(\mathbf{u} + \mathbf{u}_j) - P(\mathbf{u}_j) P^*(\mathbf{u} - \mathbf{u}_j)] d^2 \mathbf{u}_j}{\iint S_l(\mathbf{u}_j) |P(\mathbf{u}_j)|^2 d^2 \mathbf{u}_j}$$

 PTF(u) 相位传递函数

 S(u) 照明函数 P(u) 光瞳函数



极坐标相位传递函**数**建模



传递响应

基于最优照明**的差分相**衬定**量相位成像**





传递函数剖线对比





基于最优照明**的差分相**衬定**量相位成像**







基于最优照明**的差分相**衬定**量相位成像**

Hela细胞5小时动态定量相位结果(每幅相位由4幅采集图像恢复)





基于彩色复用照明的单帧差分相衬定量相位成像

复**用**照明函**数分布**

 $S_{r}(\rho,\theta) = \delta(\rho - NA_{obj})\sin(\theta + \theta_{r})$ $S_{g}(\rho,\theta) = \delta(\rho - NA_{obj})\sin(\theta + \theta_{g})$ $S_{b}(\rho,\theta) = \delta(\rho - NA_{obj})\sin(\theta + \theta_{b})$ $(\theta_{r} = \theta_{g} - 120^{\circ}, \theta_{b} = \theta_{g} + 120^{\circ})$







基于彩色复用照明的单帧差分相衬定量相位成像

Hela细胞体外动态定量相位成像(相机帧率30Hz)



MCQ-CM

Multimode Quantitative Phase Computational Microscopy



- 🗗 🗙 0 . 0 -1 □ × 文件编辑视图 模式设置 设备 差分相衬 设置 Ø X 视图0: Camera Y || 日 / [] 🖗 BI 82 % 0 5 ▼ 光源 229 ms 21 ▼ 相机指令 -11-0 5 ш ▼ 相机设置 Color 1 分辨率: RGB32 (1280x960) 前東廠 マロトと直 白平街 ne Pusi 自动: 像素尺寸 3750 nm - 肥米 1/69s () Start 懐然 ▼ 镜头设置 4X物镜: 10X物镜: ~ 40X物境: 100X物稿

模式

▼ 设置 延迟:

外径:

内径:

色彩:

▼方向

▼ 控制

MMC-CM frees the user's hands

到"设置"以激活い

載用

Flexible control and display

Questions: 定量相位成像方法的分类 定量相位成像方法的优势和劣势